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the fish prior to beginning the treatment with F. O. H. If the general condition of the fish is such that it is not possible to keep them in the laboratory for a long period of time, the following method is suggested:

The fish should be kept in a tank of fresh water and fed with a diet consisting of fresh green food and a small amount of fish meal. The water should be changed daily and the fish should be kept in a clean and bright environment. At the end of the period of treatment, the fish should be kept in a clean and bright environment and fed with a diet consisting of fresh green food and a small amount of fish meal.

Excellent preparations have been made from the material obtained in the laboratory aquaria for as long a period as three weeks. Many of these preparations have been sent to various institutions.

J. E. MURPHY

EXPERIMENTAL INVESTIGATION OF PARASITIC DEVELOPMENT IN PHYSIOLOGICAL AND BIOLOGICAL SOLUTIONS

Experiments completed at the U. S. Bureau of Fisheries, a station at Fairport, Iowa, by us this summer. Various nutrient solutions were prepared in which the trochophore of the freshwater mussel, *Anodonta imbecilis* Smith (known as the Crooked or Slough Mussel), were carried through their various developmental stages from trochophore to the glochidium stage. The glochidia of *Anodonta imbecilis* are parasitic on the gills of the shinerhead pike, *Lepisosteus platostomus* Rafinesque, for a period varying from two to several weeks, during which time the glochidia undergo marked internal changes and differentiations and emerge from their cysts at the end of this period as the fish as free-living juvenile muskells. The nutrient fluid was perfected so that this period of parasitic life on the fish could be replaced by a period *in vitro*, during which the growth and differentiations ordinarily made by the glochidium in the cyst could be studied and controlled.

The glochidia used in these first series of experiments were dissected out of their cysts on the gills of artificially infected, one, eighteen and ninety-six hours after attachment was begun. The freed glochidia were transferred to one of the solutions in which their development was to be followed. Glochidia removed from the cysts eight or ten hours after attachment to the fish gill did not attach at all in appearance from ripe glochidia in the maternal marsupium. Glochidia removed at the end of ninety-six hours showed considerable development of the organ anlagen, although the glochidia were still in a very embryonic stage, as was evidenced by the presence of a large portion of the trochophore cell mass. In the nutrient solution tested the glochidia were carried through the trochophore stage in the solution, at which time their development was controlled. Glochidia which had not attached on the fish and were just ready to emerge from the cysts. When their stage was reached in the nutrient solution the trochophore stage was reached in the nutrient solution.

For the purpose of this study, a method of preparing the nutrient solutions was better than the method of preparing the nutrient solutions. For the purpose of this study, a method of preparing the nutrient solutions was better than the method of preparing the nutrient solutions.

The nutrient solution is composed of equal parts of 1% yeast solution, 1% yeast solution, alcohol and 1% yeast solution. This nutrient solution was devised and used by Mr. J. C. Taylor in the preparation of certain nutrient solutions in the laboratory. This mixture is prepared and used in the laboratory. Place the Hydra in a clean glass jar and cover it with a layer of water. Add a small amount of water to the jar and cover it with a layer of water. Add a small amount of water to the jar and cover it with a layer of water. Add a small amount of water to the jar and cover it with a layer of water. Add a small amount of water to the jar and cover it with a layer of water.

under the microscope, but transformation in the laboratory could not be observed. The transformation of the glochidia into the adult form *in vitro* not only fails to occur, but glochidia taken from the fish, but then kept in a solution of oxygenated water in comparison with the glochidia taken with the development of the larval form, do not develop.

Several mussels which transformed in these artificial conditions were kept in river water for three weeks. The transformation without the loss of vitality of the adult, the invivile mussels making a complete growth of both shell and soft parts during that time, are reasons in every way to be very optimistic. This was, however, surprising, as there is known to be a rather high mortality, on the contrary, among mussels during the first few days after leaving the fish following the natural parasitic cycle.

The recent studies of glochidia carried in the various natural conditions *in vitro* showed that paracellulose on the surface is essential to development and for a formation if the proper food substances be supplied in the proper concentration; that the glochidium requires by its environment a much-needed protection against certain bacterial and protozoan enemies; and that the glochidium is a true parasite while on the fish, receiving all its food substances from the host fish. This last statement was repeatedly tested in a variety of experiments, and the so-called protective physiological solution containing only inorganic salts was neither adequate to produce growth and differentiation nor to maintain glochidia already well started on their way to transformation.

The successful solutions contained sodium chloride, potassium chloride, calcium chloride, sodium bicarbonate, dextrose and a mixture of amino-acids, together with small quantities of phosphates and traces of magnesium salts. Detailed data of these experiments as well as experiments on glochidia taken directly from the natural environment are to be published.

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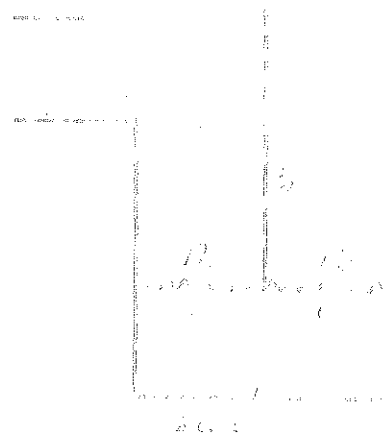
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CRITICAL POTENTIAL MEASUREMENTS: A CORRECTION FOR HIGH-EMISSION CURRENTS

In experiments on critical potentials the voltage E_c (Fig. 1) applied to a filament and grid is usually measured by means of the potentiometer P .

When the current I_c is zero then

$$E_c = \frac{E_0}{P + E_0} \quad (1)$$



However, if the current I_c is large then

$$E_c = \frac{E_0}{P + E_0} [E_0 + I_c R] \quad (2)$$

This equation is derived from Kirchhoff's law and Ohm's law

$$E = IR + I_r R \quad (3)$$

and Ohm's law

$$E = I R \quad \text{and} \quad E = I R \quad (4)$$

Combining equations (3) and (4) and eliminating I , equation (2) is obtained. In the case of networks having been applied to such a circuit, no special assumption has been necessary regarding the resistance of the filament-grid space.

Equation (2) has been tested with a one-electrode tube containing mercury. The following table shows clearly that the correction is necessary and that it becomes large

CRITICAL POTENTIAL OF MERCURY AT 0.5 MM (P. 100)

I_c microamperes	Experimentally observed	Corrected
24	7.101	7.101
160	52.0	7.124
800	232.0	73.4
Average		73.3

A further correction for initial electron velocity and contact potential has to be applied to the results.

In another test a factor of 1000 was used for the filament with a filament of 1000 ohms and a grid of 100 ohms. (2) is correct.

It can be shown that the correction is not necessary for a filament of 1000 ohms and a grid of 100 ohms.

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